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Also provided are methods for treating a disease by degrading the function of a target protein, comprising introducing, into a cell, a chimeric protein comprising a target-protein binding domain operatively linked to a protein-degradation binding domain of a protein member of the ubiquitin-mediated protein-degradation family. For example, for a variety of proteins which, when expressed in overabundant or mutated form (e.g., an oncprotein such as ras, or a genetic mutation, such as in the CF gene (cystic fibrosis gene) result in a known pathology, the chimeric protein of the invention may be used to therapeutically treat the disease, by way of reducing or completely eliminating, via protein degradation, the pathology causing protein.

This treatment comprises fusion of a protein domain which binds the target pathology causing protein (i.e., the protein which causes the illness) with a particular protein-degradation binding domain as described herein. This chimeric protein may then be delivered to the location of the protein which causes the illness by intravenous therapy or gene therapy employing the methods described herein, or any other method well-known to one skilled in the art for delivering a protein to its binding target. As used herein, "treatment of a disease" refers to a reduction in the effects of the disease, including reducing the symptoms of the disease.

In accordance with another embodiment of the present invention, there are provided methods for diagnosing cancer, said method comprising:

detecting, in said subject, a defective sequence or mutant of SEQ ID NOS:1, 3, 5, 7, 9, 11 and 13.

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In accordance with another embodiment of the present invention, there are provided diagnostic systems, preferably in kit form, comprising at least one invention nucleic acid in a suitable packaging material. The 5 diagnostic nucleic acids are derived from the SMDP and/or SCP-encoding nucleic acids described herein. In one embodiment, for example, the diagnostic nucleic acids are derived from any of SEQ ID NOs:1, 3, 5, 7, 9, 11 and 13. Invention diagnostic systems are useful for 10 assaying for the presence or absence of nucleic acid encoding SMDP and/or SCP in either genomic DNA or in transcribed nucleic acid (such as mRNA or cDNA) encoding SMDP and/or SCP.

A suitable diagnostic system includes at least 15 one invention nucleic acid, preferably two or more invention nucleic acids, as a separately packaged chemical reagent(s) in an amount sufficient for at least 20 one assay. Instructions for use of the packaged reagent are also typically included. Those of skill in the art 25 can readily incorporate invention nucleic probes and/or primers into kit form in combination with appropriate buffers and solutions for the practice of the invention methods as described herein.

As employed herein, the phrase "packaging 40 material" refers to one or more physical structures used to house the contents of the kit, such as invention 25 nucleic acid probes or primers, and the like. The packaging material is constructed by well known methods, 45 preferably to provide a sterile, contaminant-free 30 environment. The packaging material has a label which indicates that the invention nucleic acids can be used 50 for detecting a particular sequence encoding SMDP and/or SCP including the nucleotide sequences set forth in SEQ

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10 NOs:1, 3, 5, 7, 9, 11 and 13 or mutations or deletions therein, thereby diagnosing the presence of, or a predisposition for, cancer. In addition, the packaging material contains instructions indicating how the materials within the kit are employed both to detect a particular sequence and diagnose the presence of, or a predisposition for, cancer.

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The packaging materials employed herein in relation to diagnostic systems are those customarily utilized in nucleic acid-based diagnostic systems. As used herein, the term "package" refers to a solid matrix or material such as glass, plastic, paper, foil, and the like, capable of holding within fixed limits an isolated nucleic acid, oligonucleotide, or primer of the present invention. Thus, for example, a package can be a glass vial used to contain milligram quantities of a contemplated nucleic acid, oligonucleotide or primer, or it can be a microtiter plate well to which microgram quantities of a contemplated nucleic acid probe have been operatively affixed.

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"Instructions for use" typically include a tangible expression describing the reagent concentration or at least one assay method parameter, such as the relative amounts of reagent and sample to be admixed, maintenance time periods for reagent/sample admixtures, temperature, buffer conditions, and the like.

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All U.S. patents and all publications mentioned herein are incorporated in their entirety by reference thereto. The invention will now be described in greater detail by reference to the following non-limiting examples.

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**EXAMPLES**

Unless otherwise stated, the present invention was performed using standard procedures, as described, for example in Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, USA (1982); Sambrook et al., Molecular Cloning: A Laboratory Manual (2 ed.), Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, USA (1989); Davis et al., Basic Methods in Molecular Biology, Elsevier Science Publishing, Inc., New York, USA (1986); or Methods in Enzymology: Guide to Molecular Cloning Techniques Vol.152, S. L. Berger and R. K. Kimmel Eds., Academic Press Inc., San Diego, USA (1987).

**15 Two-hybrid assays.**

Library screening by the yeast two-hybrid method was performed herein as described (Durfee et al., 1993; Sato et al., 1995; Matsuzawa et al. 1996) using the pGilda plasmid encoding the desired amino acid region as bait, an appropriate cDNA library, and the EGY48 strain *S.cerevisiae* (MATα, trp1, ura3, his, leu2::pLxAp6-leu2). Cells were grown in either YPD medium with 1% yeast extract, 2% polypeptone, and 2% glucose, or in Burkholder's minimal medium (BMM) fortified with appropriate amino-acids as described previously (Sato et al., 1994). Transformations were performed by a LiCl method using 0.25 µg of pJG4-5-cDNA library DNA, and 5 µg of denatured salmon sperm carrier DNA. Clones that formed on Leu deficient BMM plates containing 2% galactose/ 1% raffinose were transferred to BMM plates containing leucine and 2% glucose, and filter assays were

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performed for  $\beta$ -galactosidase measurements as previously described.

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**1. Yeast two-hybrid screen of BAG-1 binding proteins to obtain cDNA encoding Siah-1 $\alpha$ .**

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The mouse BAG-1 amino acid sequence was cloned into the pGilda plasmid and used as bait to screen a human Jurkat T-cell cDNA library. From an initial screen of  $\sim 1.6 \times 10^7$  transformants, 298 clones were identified that trans-activated the LEU2 reporter gene based on ability to grow on leucine-deficient media. Of those, 30 colonies were also positive for  $\beta$ -galactosidase. These 30 candidate transformants were then cured of the LexA/BAG-1 bait plasmid by growth in media containing histidine and then mated with each of 5 different indicator strains of cells containing one of following LexA bait proteins: BAG-1 (1-219), Bax (1-171), v-Ras, Fas (191-335), or Lamin-C. The mating strain was RFY206 (MATα, his3Δ200, leu2Δ3, lys2Δ201, ura3Δ5, trp1Δ::hisG), which had been transformed with pGilda-BAG-1 or various control proteins and selected on histidine-deficient media. This resulted in 23 clones which displayed specific two-hybrid binding interactions with BAG-1. DNA sequencing analysis revealed 4 cDNAs encoding portions of Siah-1.

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**2. Isolation of full-length human Siah-1 $\alpha$  cDNAs.**

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To obtain the complete sequence of human Siah-1, cDNA fragments containing the 5' end of human Siah-1 were PCR-amplified from Jurkat randomly primed cDNAs by using a forward primer 5' 30 3GGAAATTCGGGACTTATGCGATGTAAACCA-3' (SEQ ID NO:42) containing an EcoRI site and a reverse primer 5'

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5 CACCCRAGTTGCGAATGGA-3' (SEQ ID NC:43), based on sequences  
of EST database clones (NCBI ID: AA054277, AA258606,  
AA923663, AA418482, and AI167464). The PCR products were  
10 digested with EcoRI and BamHI, then directly subcloned  
5 into the EcoRI and SalI sites of pCL plasmid into which  
the cDNA derived from pJG4-5-Siah (22-298) had previously  
been cloned, as a BamHI - Khol fragment. The complete  
15 human Siah-1α cDNA and amino acid sequence is set forth  
in SEQ ID Nos:1 and 2, respectively. The human Siah-1α  
10 sequence contains 16 N-terminal amino acids that are not  
present in the human Siah-1β protein.

20 3. Yeast two-hybrid screen of Siah-1 binding proteins to  
obtain cDNA encoding SIP-L and SIP-S.

25 Human Siah-1α cDNA encoding amino acids 22-298  
15 of SEQ ID NC:1 (corresponding to amino acids 6-202 set  
forth in Nemani et al., supra) was cloned into the pGilda  
plasmid and used as a bait to screen a human embryonic  
30 brain cDNA library (Invitrogen) in EGY48 strain  
*S.cerevisiae*. From an initial screen of ~2.0 X 10<sup>7</sup>  
20 transformants, 322 clones were identified that trans-  
activated the LEU2 reporter gene based on ability to grow  
on leucine-deficient media. Of those, 32 colonies were  
also positive for β-galactosidase. These 32 candidate  
35 transforms were then cured of the LexA/Siah-1 bait  
25 plasmid by growth in media containing histidine and then  
mated with each of 5 different indicator strains of cells  
containing one of following LexA bait proteins: Siah-  
1(22-298), Bax (1-171), v-Ras, Fas (131-335), or BAG-1.  
40 The mating strain was RPY206 which had been transformed  
30 with pGilda-Siah-1 or various control proteins and  
selected on histidine-deficient media. This resulted in  
11 clones which displayed specific two-hybrid  
50 interactions with Siah-1. DNA sequencing analysis

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revealed 5 cDNAs encoding portions of SIP-L, 1 cDNA encoding portions of SIP-S, 3 cDNAs encoding portions of APC(2681-2643), and 2 cDNAs encoding portions of Sish-1. The SIP-L and SIP-S clones were sequenced and the resulting nucleotide sequences are set forth in SEQ ID Nos:2 and 5, respectively.

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4. Yeast two-hybrid screen of Skp1 binding proteins to obtain cDNA encoding SAF-1 and SAD.

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Human Skp1 cDNA encoding amino acids 91-163 of (Zhang et al., 1995, *Cell*, 82:915-925) was cloned into the pG4dia plasmid as a bait to screen a human embryonic brain cDNA library (Invitrogen) in EGY48 strain *S.cerevisiae*. From an initial screen of ~1.2 X 10<sup>6</sup> transformants, 130 clones were identified that trans-15 activated the LEU2 reporter gene based on ability to grow on leucine-deficient media. Of those, 36 colonies were also positive for β-galactosidase. These 36 candidate 20 transformants were then cured of the LexA/BAG-1 bait plasmid by growth in media containing histidine and then mated with each of 5 different indicator strains of cells 25 containing one of following LexA bait proteins: Skp1 (91-163), SIP-1, Bax (1-171), v-Ras, Fas (191-335), or Siah-1. The mating strain was RMY206 which had been transformed with pG4dia-Skp1 or various control proteins 30 and selected on histidine-deficient media. This resulted in 3 clones which displayed specific two-hybrid interactions with Skp1 and 18 clones which displayed specific two-hybrid interactions with both Skp1 and SIP-L. DNA sequencing analysis revealed 12 cDNAs 35 encoding portions of SAF-1 and 3 cDNAs encoding portions of SAD. The SAF-1 and SAD clones were sequenced and the resulting nucleotide sequences are set forth in SEQ ID Nos:7 (SAF-1α), 9 (SAF-1β), and 13 (SAD).

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**5. Isolation of full-length SAF-2 cDNAs.**

Full-length cDNA encoding a human SAF-2 protein was PCR-amplified from ZAPII Jurkat cDNA library (Stratagene) by using a forward primer 5'-  
10 GTGAATTCTATGCAACTTGTACCTGATATAGAGTC-3' (SEQ ID NO:44)  
containing an EcoRI site and a reverse primer 5'-  
15 GGACTCGAGGCTCTACAGAGGCC-3' (SEQ ID NO:45), based on human  
DNA sequence from clone 341E18 on chromosome 6p11.2-12.3  
(ALC31178). The PCR products were digested with EcoRI  
20 and XbaI, then directly subcloned into the EcoRI and XbaI  
sites of the plasmid pCDNAJ. The corresponding plasmid  
was sequenced and the results are set forth in SEQ ID  
Nos: 11 and 12.

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**6. Yeast two-hybrid screen of SIP-L binding proteins.**

15 The human SIP-L cDNA encoding full-length SIP-L was cloned into the pG4Lda plasmid as a bait to screen a  
20 human embryonic brain cDNA library (Invitrogen) in CGY48 strain *S.cerevisiae*. From an initial screen of ~1.5 X 10<sup>6</sup> transformants, 410 clones were identified that trans-  
25 20 activated the LEU2 reporter gene based on ability to grow on leucine-deficient media. Of those, 68 colonies were also positive for  $\beta$ -galactosidase. These 32 candidate transformants were then cured of the LexA/SIP-L bait  
30 25 plasmid by growth in media containing histidine and then mated with each of 37 different indicator strains of  
cells containing one of following LexA bait proteins:  
35 SIP-L, Bax (1-171), v-Ras, Fas (191-335), or SAG-1. The mating strain was RFY20e which has been transformed with  
40 pG4Lda-SIP-L or various control proteins and selected on  
45 histidine-deficient media. This resulted in 16 clones which displayed specific two-hybrid interactions with  
50 SIP-L. DNA sequencing analysis revealed 3 cDNAs encoding

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10 portions of Skp1, 1 cDNA encoding portions of Siah-1, and  
11 cDNAs encoding portions of SIP-L. These results  
indicate that SIP-L binds to Skp1 and Siah-1 proteins,  
15 and is able to homodimerize with SIP isoforms.

15 5 7. A cell proliferation functional assay of SIP/Siah  
interaction

20 The effects of invention SIP-L and SIP-S  
proteins on Siah-1-induced cell cycle arrest in 293T  
epithelial cancer cells was examined and the results are  
10 shown in Figure 4. Human embryonic kidney 293 cells were  
maintained in high-glucose DMEM medium containing 10%  
fetal calf serum, 1 mM L-glutamine, and antibiotics.  
25 Cells (~5 x 10<sup>4</sup>) in 60 mm plates were transfected with a  
total of 3.0 µg of plasmid DNAs encoding Siah-1 alone or  
15 together with SIP or SIP-S by a calcium phosphate  
precipitation technique. After 24 hours, the cells were  
harvested and the number of viable and dead cells were  
30 counted using trypan blue dye exclusion assays.  
Efficiency of transient transfection was estimated by in  
20 situ β-galactosidase assay using a portion of the  
transfected cells. The transient transfection efficiency  
35 of the 293 cells was consistently ~90%.

40 As revealed in Figure 4, over-expression of  
Siah-1 resulted in decreased numbers of viable cells  
45 after 24 hours, without an increase in cell death. Thus,  
Siah-1 suppresses proliferation of 293 cells. Co-  
transfection of SIP-L with Siah-1 did not substantially  
alter Siah-1-mediated growth suppression. In contrast,  
50 the SIP-S protein abrogated the growth suppressive  
effects of Siah-1, which indicates that the invention  
SIP-S protein affects Siah-1 intracellularly in a  
different manner than SIP-L.

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**8. In vitro SIP:Siah-1 protein interaction assays.**

Complementary cDNA encoding SIP-L was cloned into pGEX-4T-1 and expressed in XL-1-blue cells (Stratagene, Inc.), and affinity-purified using glutathione-Sepharose as is well-known in the art. Purified GST-fusion proteins (0.5-1.0 µg immobilized on 10-20 µl of glutathione beads) and 2.5 µl of rat reticulocyte lysates (TNT-Lysates; Promega, Inc.) containing 35S-labeled in vitro translated (IVT) Siah-1 proteins were incubated in 0.1 ml of HKNEN (10 mM HEPES [pH 7.2], 142 mM KCl, 5 mM MgCl<sub>2</sub>, 2 mM EGTA, 0.1% NP-40) at 4°C for 30 minutes. The beads were washed 3X with 1 ml HKNEN solution, followed by boiling in 25 µl of Laemmli-SDS sample buffer. The eluted proteins were analyzed by SDS-PAGE (12%) and detected by fluorography. Use of equivalent amounts of intact GST-fusion proteins and successful IVT of each protein was confirmed by SDS-PAGE analysis using Coomassie staining or autoradiography, respectively.

The results are shown in Figure 5A and indicate that Siah-1 binds to SIP-L and homodimerizes in vitro.

**9. Co-immunoprecipitation Assay of SIP:Siah-1.**

Two × 10<sup>6</sup> 293T cells in 100 mm plates were transiently transfected with 10 µg of pCNA3-myc-SIP-L and 15 µg of pcDNA3-HA-Siah-1 (amino acids 97-296 of SEQ ID NO:2). Twenty-four hours later, cells were disrupted by sonication in 1 ml of HKNEN solution containing 0.2% NP-40, 0.1 µM PXSF, 5 µg/ml leupeptin, 1 µg/ml aprotinin, and 1 µg/ml pepstatin. After preclarifying with normal mouse IgG and 10 µl protein A-agarose, immunoprecipitations were performed using 10 µl of anti-

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5 myc antibody-conjugated sepharose (Santa Cruz) to  
precipitate the myc-SIP-L fusion, or an anti-IgG as a  
control at 4°C for 4 hours. After extensive washing in  
HMMEN solution, immune-complexes were analyzed by SDS-  
10 PAGE/immunoblotting using anti-MA antibody 12CA5  
(Boehringer Mannheim), followed by HRPase-conjugated goat  
anti mouse immunoglobulin (Amersham, Inc.), and detected  
15 using an enhanced chemiluminescence (ECL) system  
(Amersham, Inc.).

20 10 The results are shown in Figure 5B and indicate  
that SIP proteins bind to Siah-1 intracellularly.

25 10. Yeast two-hybrid assay of Siah-1:APC binding  
specificity.

30 15 One ug of plasmids encoding fusion proteins of  
the LexA DNA-binding domain fused to Siah-1, APC(2681-  
284), BAG-1, Bax, Ras, Fas, FLICE were co-transformed into  
yeast strain EGI40 with 1 ug of pCG4-5 plasmid encoding  
35 20 fusion proteins of the B42 trans-activation domain fused  
to APC(2681-2843) and Siah-1. Transformed cells were  
grown on semi-solid media lacking leucine or containing  
leucine as a control which resulted in equivalent amounts  
40 25 or growth for all transformants. Plasmid combinations  
that resulted in growth on leucine-deficient media within  
4 days were scored as positive (+).  $\beta$ -galactosidase  
activity of each colony was tested by filter assay and  
scored as blue (+) versus white (-) after 60 minutes.

45 30 The results are shown in Table 1, and indicate  
that APC interacts specifically by direct binding with  
Siah-1, and not with BAG-1, Bax, Ras, Fas nor FLICE.

50 55 Table 1: Specific Interaction of Siah with SIP

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Lex A	B42	Leu <sup>+</sup>	$\beta$ -Gal <sup>+</sup>
Siah-1	APC (2681-2843)	+	+
APC (2681-2843)	Siah-1	+	+
BAG-1	APC (2681-2843)	-	-
Bax	APC (2681-2843)	-	-
Ras	APC (2681-2843)	-	-
Fas	APC (2681-2843)	-	-
FLICE	APC (2681-2843)	-	-
empty	APC (2681-2843)	-	-

11. Yeast two-hybrid assay of Siah-1:SIP binding specificity.

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15 One  $\mu$ g of plasmids encoding fusion proteins of the LexA DNA-binding domain fused to Siah-1, Siah-2, BAG-1, Bax, Ras, Fas, FLICE, and SIP-L were co-transformed into yeast strain EGY48 with 1  $\mu$ g of pJG4-5 plasmid encoding fusion proteins of the B42 trans-activation domain fused to SIP-L, SIP-S, Siah-1, Siah-2, BAG-1, Bax, and Ras. Transformed cells were grown on semi-solid media lacking leucine or containing leucine as a control which resulted in equivalent amounts of growth for all transformants. Plasmid combinations that resulted in 20 growth on leucine-deficient media within 4 days were 25 scored as positive (+).  $\beta$ -galactosidase activity of each

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colony was tested by filter assay and scored as blue (+) versus white (-) after 60 minutes.

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The results are shown in Table 2, and indicate that SIF proteins interact specifically by direct binding with Siah proteins. SIP-L was found to interact with Siah-1 and Siah-2, and not with BAC-1, Bax, Ras, Fas nor FLICE. SIP-S was also found to interact with Siah-1. Table G also reveals that the SIP-L homodimerization domain is within amino acids 73-228 of SIP-L (SINQ ID 10 NO:4)

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## Specific Interaction of Siah with SIP

Table 2

Lex A	B42	Leu <sup>+</sup>	$\beta$ -Gal <sup>+</sup>
Siah-1	SIP-L	+	+
Siah-1	SIP-S	+	+
Siah-2	SIP-L	+	+
BAG-1	SIP-L	-	-
Bax	SIP-L	-	-
Ras	SIP-L	-	-
FLT4CE	SIP-L	-	-
empty	SIP-L	-	-
SIP-L	Siah-1	-	+
SIP-L	Siah-2	+	+
SIP-L	BAG-1	-	-
SIP-L	Bax	-	-
SIP-L	Ras	-	-
SIP-L	SIP-L	+	+
SIP-L	SIP-S	-	-

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## 12. Mapping of Siah-APC interaction domains.

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Expression plasmids encoding fusion proteins of Siah-1 $\alpha$  fragments corresponding to: SEQ ID NO:2 amino acids 29-298; 22-251; 22-193; 97-298; and 46-102, fused to the B-42 trans-activation domain were co-transformed into yeast EGY48 cells with a plasmid encoding a chimeric fusion protein of the Lex A DNA-binding domain fused to amino acids 2681-2843 of APC "APC(2681-2843)." Transformed cells were grown on semi-sciid media lacking leucine or containing leucine as a control. Plasmid combinations that resulted in growth on leucine-deficient media within 4 days were scored as positive (+).  $\beta$ -

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galactosidase activity for each colony was tested by filter assay and scored as blue (+) versus white (-) ( $\beta$ -gal) based on a 1 hour of color development.

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The results are shown in Figure 3 and indicate that a region within the 47 carboxy terminal amino acids of Siah-1 $\alpha$  (SEQ ID NO:7) is required for binding to AFC.

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**13. Mapping of SKP-1, SIP-1, SAF-1, and SAD interaction domains.**

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Expression plasmids encoding fusion proteins of SAF-1 $\alpha$  and functional fragments thereof corresponding to SEQ ID NO:8 amino acids 68-443; 80-443; and 258-443, were fused to the B-42 trans-activation domain. Likewise, expression plasmids encoding fusion proteins of SAD and functional fragments thereof corresponding to SEQ ID NO:14 amino acids 128-447; and 360-447, were fused to the B-42 trans-activation domain. These SAF-1-fragment- and SAD-fragment-B-42 fusion proteins were co-transformed into yeast EGY48 cells with a plasmid encoding a chimeric fusion protein of the Lex A DNA-binding domain fused to either SKP1, SIP-1, SNF1, or SAD. Transformed cells were grown on semi-solid media lacking leucine or containing leucine as a control. Plasmid combinations that resulted in growth on leucine-deficient media within 4 days were scored as positive (+).  $\beta$ -galactosidase activity for each colony was tested by filter assay and scored as blue (+) versus white (-) ( $\beta$ -gal) based on a 1 hour of color development.

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The results are shown in Figure 6A and 6B. Figure 6A indicates that SAF-1 interacts by direct binding to Skp1, SIP-1 and SAD, but does not interact with Siah-1. A region within the SAF-1 fragment

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corresponding to amino acids 60-257 of SEQ ID NO:J is required for SIP-L interaction, whereas a region within amino acids 258-443 of SAF-1 is required for Skp1 and SAD interaction.

Figure 6B indicates that SAD interacts by direct binding to Skp1, SIP-L and SAF-1, but does not interact with Siah-1. A region within the SAD fragment corresponding to amino acids 1-127 of SEQ ID NO:14 is required for SAF-1 interaction; a region within amino acids 128-359 of SAD is required for Skp1 interaction; and a region within amino acids 360-447 of SEQ ID NO:14 is required for SIP-L interaction.

14. Effect of Siah-1 over-expression on stability of  
β-catenin.

293T cells were transiently transfected with a plasmid encoding myc-tagged β-catenin and either pcDNA3, pcDNA3-Siah-1, or pcDNA3-Siah-1(97-298); amino acids 97-298 of SEQ ID NO:2. Whole cell lysates were prepared, normalized for total protein content (25 pg per lane) and analyzed by SDS-PAGE/immunoblotting using an anti-Myc tag antibody.

Figure 7 indicates that expression of full-length Siah-1 abolishes, by degradation, the presence of β-catenin within cells, whereas expression of amino acids 97-298 of Siah-1 (SEQ ID NO:2) does not result in β-catenin degradation. Thus, a region within amino acids 1-96 of SEQ ID NO:2 (Siah-1<sub>u</sub>), which contains the N-terminal "Ring" domain, is required for protein degradation.

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5                   **15. Demonstration of SIP-mediated degradation of a target protein, TRAF6.**

10                   An invention SIP-based method for targeted degradation of proteins was applied to the degradation  
5                   of TRAF proteins. The schematic in Figure 9 shows the strategy employed for targeted degradation of specific  
15                  TRAF-family proteins. A chimeric protein is expressed from the plasmid pcDNA3 in which SIP-L (SEQ ID NO:3) is fused with bacterial thioredoxin containing various TRAF-  
20                  binding peptides displayed on the surface of thioredoxin, as described by Brent and colleagues (Celsus, et al.  
Nature, **380**: 548, 1996; Cohen, et al. Proc. Natl. Acad. Sci., **95**: 14272, 1998; Geyer, et al. Proc. Natl. Acad. Sci., **96**: 8562, 1999; Fabrizio, et al. Oncogene, **18**: 4357, 1999). The TRAF-binding peptide binds to a member  
25                  of the TRAF-family, and targets the TRAF-protein for ubiquitination and subsequent proteasome-dependent  
30                  degradation because the SIP-region of the chimeric protein recruits ubiquitin-conjugating enzymes (E2s) to  
20                  the protein complex.

35                   Isolation of target-protein binding domain peptides that selectively bind TRAF2 and TRAF6.

40                   A peptide aptamer library was screened by the yeast two-hybrid method to identify peptides that bind to  
45                  either TRAF2 or TRAF6 using the methods coscribed in Ito, et al. J Biol Chem, **274**:22414, 1999. TRAFs are a family  
50                  of signal transducing proteins involved in cytokine receptor signaling inside cells. The sequences of the resulting TRAF-binding peptides are set forth in (Tables  
3 and 4).

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**TABLE 3****Selected Traf 1 Aptamer Clones**

	<b>Clones</b>	<b>(SEQ_ID NCBI)</b>	<b>SLXCLLR motif</b>
10	219	(15)	SESPGALRSGSLLRISLSEIC
5	230	(16)	VRGRIRSGSLLRISLSEIC
	221	(17)	LLRIGCIPLLMLRBRGVVFPL
15	208	(18)	VLFLSLRLFWGNIINTVVMGRLL
	215	(19)	CRSIGVIVGGTEAAGAPTFI
			<b>LS motif</b>
10	208	(20)	VLFLSLRLFWGNIINTVVMGRLL
	213	(21)	WLBRROLVGWFPLLSKVMVGI
20	218	(22)	SLGLSVCIGRRAGGGFRGFEG
	237	(23)	RPA <sup>*</sup> SIGVCVVVRVGICLGM
			<b>LV motif</b>
15	209	(24)	SAV <sup>*</sup> LVYVVAALRGRGFPI
	227	(25)	HGGGRGA <sup>*</sup> MSVMM <sup>*</sup> CGIFRL
25			<b>Non-Consensus motif</b>
	231	(26)	RGRVIGMWVGRLRCRMFLV

**TABLE 4****Selected Traf 6 Aptamer Clones**

	<b>Clones</b>	<b>(SEQ_ID NCBI)</b>	<b>WS motif</b>
30	625	(27)	VDWAVYSVVVEYTT*
	631	(28)	KTSVILVWRLSLFFCLYRLSL*
35	606	(29)	ANRCME*
	25	628	EGTILSKRMZBTHN*
	640	(31)	SZDNTICSGM*
	604	(32)	DVPMCARACARO*
	607	(33)	LERVVARZVL*
40	602	(34)	VADVUVFKGYVF*
	30	602	<b>DWxVF motif</b>
	613	(35)	VADVUVENGYYVF*
			SDWGVPE*
45	603	(36)	<b>Non-Consensus motif</b>
	35	609	PEKKLEGRKYCLXK*
	612	(38)	LLYGALA*
	616	(39)	GAIKFHAESES*
	632	(40)	PMAMD*
	639	(41)	CESEM*
50	40	*	ISVWHGIGSDSD*
			* Termination codon

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SIP-fusion Chimeric protein construction

An invention SIP-fusion chimeric construct is generated by combining the open reading frame (ORF) of SIP, followed immediately by restriction enzyme sites allowing for subcloning of desired target-protein-binding domains (e.g. peptides or protein domains). These SIP-fusions are then transfected into mammalian cells to eliminate by protein degradation specific target proteins which bind the subcloned peptides/protein domains by recruiting them into the ubiquitin conjugating complex.

The parent SIP-vector (SIPpcDNA3.1) cassette was constructed as follows:

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Oligonucleotides corresponding to the 5' and 3' end of SIP, were used in PCR to amplify the entire ORF of SIP, (SEQ ID NO:3). The forward primer contains a *Hind III* restriction site linker (5'-GATCAAAGCTTACGGCTTCAGAAGAGCTACAG; (SEQ ID NO:46) restriction site is underlined) followed immediately by the SIP, (SEQ ID NO:3) start codon; the reverse primer 30 contains an *EcoRI* restriction site and mutations in the stop codon allowing for translational readthrough (5'-GATCGAATTCTccAANTCAGTGCTCCTTTGGCTTG; (SEQ ID NO:47) mutated stop codon is in lowercase). The generated PCR product was then agarose gel-purified and digested with 35 *Hind III* and *EcoRI* restriction enzymes (New England Biolabs; Beverly, MA). The product was again gel-purified before ligating into *Hind III/EcoRI* digested pcDNA3.1 expression vector (Invitrogen; Carlsbad, CA) with T4-DNA ligase (New England Biolabs). This construct was termed 40 SIPpcDNA3.1.

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For the construction of SIP-thioredoxin (Trx) peptide-aptamer fusions, clones from a peptide-aptamer library screened against Traf6 (see Table 4) were amplified by PCR with the following primers:  
5 Forward: 5'-CCCTGAAUCCAAATGAGCGATAAATTTCACC (SEQ ID NO:48) EcoRI underlined; Reverse: 5'-  
10 CATCTCGACTAGATGCCAGCTAGGCCAGTTA (SEQ ID NO:49) Xba I underlined.

The resulting PCR products (~350-370bp) contain  
15 the NPF of thioredoxin (Trx) with the selected peptide  
aptamers inserted into its active-loop. The products  
were then digested with *Eco*RI and *Xba* I before ligating  
20 into the *Eco*RI/*Xba*I-digested SIPpcDNA3.1 cassette using  
*T4*-DNA ligase. Final clone constructs were numbered and  
25 confirmed by sequencing before using in transfection  
studies.

#### Transfection

30 HEK293T cells were transiently transfected by a  
lipofectamine method with various amounts (1 vs 4 µg) of  
35 pcDNA3 plasmids encoding either SIP:TR fusion protein  
lacking a TRAF6-binding peptide ("SIP") or SIP:TR fusion  
protein displaying one of the peptides shown in Table 4  
above (set forth in Figure 10 as S603, S604, S606). In  
40 some cases, the proteasome inhibitor MG132 (10 µM) was  
25 added to cultures to prevent protein turnover. SIP+ in  
Figure 10 corresponds to the control expression product  
of parental construct SIP pcDNA3.1

45 To determine the efficacy of the SIP:TRAF-  
binding peptide chimeric proteins, levels of TRAF6  
50 protein were then measured two days later by  
immunoblotting using a anti-TRAF6-specific antiserum

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(Santa Cruz Biotech, Inc.) in experiments where HEK293T cell lysates were normalized for total protein content (25  $\mu$ g per lane). The cell lysates were analyzed by SDS-PAGE/immunoblotting using an enhanced chemiluminescence detection method, as described previously (Leu, et al. J Biol Chem, 274: 22414, 1999). The results shown in the left panel of Figure 10 show that SIP:TR fusion proteins displaying TRAF6-binding peptides (S603, S604, and S613) induce a reduction in TRAF6 protein levels, with the S603 peptide representing the most potent of these.

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To determine the specificity of the SIP:TRAF-binding peptide chimeric proteins, the same immunoblots were reprobed with an antiserum against SIP to demonstrate equivalent levels of production of SIP:TR fusion proteins, or with antibodies specific for TRAF2 to reveal selective degradation of TRAF6 but not TRAF2. The results shown in the right panel of Figure 10 show that addition of a proteasome inhibitor, MG132, prevents the reductions in TRAF6. Note also that TRAF2 protein is not degraded, demonstrating the specificity of the targeting approach.

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While the invention has been described in detail with reference to certain preferred embodiments thereof, it will be understood that modifications and variations are within the spirit and scope of that which is described and claimed.

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Summary of Sequences

10 SEQ ID NO:1 is a cDNA (and the deduced amino acid sequence) encoding a Siah 1 $\alpha$  of the present invention.

15 SEQ ID NO:2 is the deduced amino acid sequence of a  
5 Siah 1 $\alpha$  protein of the present invention encoded by  
SEQ ID NO:1.

20 SEQ ID NO:3 is a cDNA (and the deduced amino acid sequence) encoding a human SIP-L polypeptide of the present invention.

10 SEQ ID NO:4 is the deduced amino acid sequence of a  
25 human SIP-L protein of the present invention encoded by  
SEQ ID NO:3.

30 SEQ ID NO:5 is a cDNA (and the deduced amino acid sequence) encoding a human SIP-S polypeptide of the  
15 present invention.

35 SEQ ID NO:6 is the deduced amino acid sequence of a  
human SIP-S protein of the present invention encoded by  
SEQ ID NO:5.

40 SEQ ID NO:7 is a cDNA (and the deduced amino acid sequence) encoding a human SAF-1 $\alpha$  polypeptide of the  
25 present invention.

45 SEQ ID NO:8 is the deduced amino acid sequence of a  
SAF-1 $\alpha$  protein of the present invention encoded by  
SEQ ID NO:7.

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SEQ ID NO:9 is a cDNA (and the deduced amino acid sequence) encoding a human SAD-13 polypeptide of the present invention.

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SEQ ID NO:10 is the deduced amino acid sequence of a SAD-13 protein encoded by SEQ ID NO:9.

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SEQ ID NO:11 is a cDNA (and the deduced amino acid sequence) encoding a human SAD-2 polypeptide of the present invention.

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SEQ ID NO:12 is the deduced amino acid sequence of a SAD-2 protein encoded by SEQ ID NO:11.

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SEQ ID NO:13 is a cDNA (and the deduced amino acid sequence) encoding a human SAD polypeptide of the present invention.

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SEQ ID NO:14 is the deduced amino acid sequence of a SAD protein encoded by SEQ ID NO:13.

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Claims

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That which is claimed is:

- 10 1. Isolated nucleic acid encoding a Sish-Mediated-Degradation-Protein (SMDP) and/or SFQ-Complex-Protein (SCP), or a functional fragment thereof.
- 15 5 2. Isolated nucleic acid encoding Sish-Mediated-Degradation-Protein (SMDP) and/or SFQ-Complex-Protein (SCP), or functional fragments thereof, selected from:
  - (a) DNA encoding the amino acid sequence set forth in SEQ ID Nos:2, 4, 6, 8, 10, 12 or 14, or
  - (b) DNA that hybridizes to the DNA of (a) under moderately stringent conditions, wherein said DNA encodes biologically active SMDP and/or SCP, or
  - (c) DNA degenerate with respect to either (a) or (b) above, wherein said DNA encodes biologically active SMDP and/or SCP.
- 20 10 30 3. A nucleic acid according to claim 2, wherein said nucleic acid hybridizes under high stringency conditions to the SMDP and/or SCP coding portion of any of SEQ ID NOS:1, 3, 5, 7, 9, 11 and 13.
- 25 15 35 40 45 50 4. A nucleic acid according to claim 2, wherein the nucleotide sequence of said nucleic acid is substantially the same as set forth in any of SEQ ID NO:1, 3, 5, 7, 9, 11 and 13.
5. A nucleic acid according to claim 2, wherein the nucleotide sequence of said nucleic acid is the same as that set forth in any of SEQ ID NOS:1, 3, 5, 7, 9, 11 and 13.
6. A nucleic acid according to claim 2, wherein said nucleic acid is cDNA.

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5           7. A vector containing the nucleic acid of claim  
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10           8. Recombinant cells containing the nucleic acid  
of claim 2.

15           9. An oligonucleotide comprising at least 15  
nucleotides capable of specifically hybridizing with a  
the nucleotide sequence set forth in any of SEQ ID Nos:1,  
3, 5, 7, 9, 11 and 13.

20           10. An oligonucleotide according to claim 9,  
wherein said oligonucleotide is labeled with a detectable  
marker.

25           11. An antisense-nucleic acid capable of  
specifically binding to mRNA encoded by said nucleic acid  
according to claim 2.

30           15       12. A kit for detecting the presence of the SMDP  
and/or SCP cDNA sequence comprising at least one  
oligonucleotide according to claim 10.

35           19. An isolated Siah-Mediated-Degradation-Protein  
(SMDP) and/or SFC-Complex-Protein (SCP) characterized by  
20       having ability to bind to at least one SMDP and/or SCP.

40           14. A SMDP and/or SCP according to claim 13,  
wherein the amino acid sequence of said protein comprises  
substantially the same sequence as any of SEQ ID Nos:2,  
4, 6, 8, 10, 12 or 14.

45           25       15. A SMDP and/or SCP according to claim 14  
comprising the same amino acid sequence as set forth in  
any of SEQ ID Nos:2, 4, 6, 8, 10, 12 or 14.

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16. A SMDP and/or SCP according to claim 13,  
wherein said protein is encoded by a nucleotide sequence  
comprising substantially the same nucleotide sequence as  
set forth in SEQ ID Nos:1, 3, 5, 7, 9, 11 or 13.

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17. A SMDP and/or SCP according to claim 16,  
wherein said protein is encoded by a nucleotide sequence  
comprising the same sequence as set forth in SEQ ID  
Nos:1, 3, 5, 7, 9, 11 or 13.

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18. A method for expression of a SMDP and/or SCP  
protein, said method comprising culturing cells of claim  
6 under conditions suitable for expression of said SMDP  
and/or SCP.

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19. An isolated anti-SMDP and/or SCP antibody  
having specific reactivity with a SMDP and/or SCP  
according to claim 13.

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20. Antibody according to claim 19, wherein said  
antibody is a monoclonal antibody.

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21. An antibody according to claim 20, wherein said  
antibody is a polyclonal antibody.

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22. A composition comprising an amount of the  
antisense-nucleic acid according to claim 11 effective to  
inhibit expression of a human SMDP and/or SCP and an  
acceptable hydrophobic carrier capable of passing through  
a cell membrane.

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23. A transgenic nonhuman mammal expressing  
exogenous nucleic acid encoding a SMDP and/or SCP.

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24. A transgenic nonhuman mammal according to claim

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5            23, wherein said nucleic acid encoding said SMDP and/or SCP has been mutated, and wherein the SMDP and/or SCP so expressed is not native SMDP and/or SCP.

10            25. A transgenic nonhuman mammal according to claim  
5        23, wherein the transgenic nonhuman mammal is a mouse.

15            26. A method for identifying nucleic acids encoding  
a mammalian SMDP and/or SCP, said method comprising:  
            contacting a sample containing nucleic acids with an  
20        oligonucleotide according to claim 9, wherein said  
10        contacting is effected under high stringency  
hybridization conditions, and identifying compounds which  
hybridize thereto.

25            27. A method for detecting the presence of a human  
SMDP and/or SCP in a sample, said method comprising  
15        contacting a test sample with an antibody according to  
claim 19, detecting the presence of an antibody-SMDP  
and/or SCP complex, and therefor detecting the presence  
30        of a human SMDP and/or SCP in said test sample.

35            28. Single strand DNA primers for amplification of  
20        SMDP and/or SCP nucleic acid, wherein said primers  
comprise a nucleic acid sequence derived from the nucleic  
acid sequences set forth as SEQ ID NOs:1, 3, 5, 7, 9, 11  
40        and 13.

45            29. A method for modulating the activity of an  
25        oncogenic protein, comprising contacting said oncogenic  
proteins with a substantially pure SMDP and/or SCP, or a  
oncogenic protein-binding fragment thereof.

50            30. A bioassay for evaluating whether test  
compounds are capable of acting as agonists or

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antagonists for SMDP and/or SCP proteins, or functional fragments thereof, wherein said bioassay comprises:

- 10                         (a) culturing cells containing:  
5                             DNA which expresses an SMCP and/or  
                           SCP or functional fragments thereof,  
15                             wherein said culturing is carried out in the  
                           presence of at least one compound whose ability  
                           to modulate an activity of an SMDP and/or SCP  
                           is sought to be determined, wherein said  
20                             activity is selected from a protein:protein  
                           binding activity or a protein degradation  
                           activity and thereafter  
25                         (b) monitoring said cells for either an increase or  
                           decrease in the level of protein:protein  
                           binding or protein degradation.

30                         31. A method for modulating an activity mediated by  
                           a SMDP and/or SCP protein, said method comprising:  
                           contacting said SMDP and/or SCP protein with an  
                           effective, modulating amount of said agonist or  
                           20 antagonist identified by claim 30.

35                         32. The method of claim 31, wherein said modulated  
                           activity is the binding of Siah-1 to APC.

40                         33. A method for modulating the protein degradation  
                           activity mediated by an SMDP and/or SCP protein, said  
                           method comprising:  
45                             contacting said SMDP and/or SCP protein with an  
                           effective, modulating amount of said agonist or  
                           antagonist identified by claim 30.

50                         34. A therapeutic composition comprising a compound  
                           30 selected from an SMDP and/or SCP, or functional fragment

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thereof, a SMDP and/or SCP modulating compound identified according to claim 30, or an anti-SMDP and/or SCP antibody; and a pharmaceutically acceptable carrier.

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10 35. A method of treating a pathology characterized by abnormal cell proliferation or abnormal inflammation, said method comprising administering an effective amount of the composition according to claim 34.

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20 36. A method of inducing the degradation of the function of a target protein, said method comprising: expressing, in a cell, a chimeric protein comprising a target-protein binding domain operatively linked to a protein-degradation binding domain of a protein member of the ubiquitin-mediated protein-degradation family.

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30 37. A method of determining the function of a target protein, said method comprising: expressing, in a first cell, a chimeric protein comprising a target-protein binding domain operatively linked to a protein-degradation binding domain of a protein member of the ubiquitin-mediated protein-degradation family; and comparing the phenotype of said first cell to the phenotype of a control second cell.

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40 38. A method of identifying a nucleic acid molecule encoding a protein that modulates a cellular phenotype, said method comprising:  
45 (a) expressing, in a cell, a chimeric nucleic acid comprising a member of a nucleic acid library fused to nucleic acid encoding a protein degradation binding domain of a protein member of the ubiquitin-mediated protein degradation family; and

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(b) screening said cells for a modulation of  
said phenotype.

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38. The method of claim 36, wherein the phenotype  
is selected from the group consisting of: cell  
5 proliferation, cell survival, cell death, cell secretion,  
and cell migration.

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40. A chimeric nucleic acid identified according to  
claim 36.

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41. A nucleic acid library comprising a plurality  
10 of chimeric nucleic acids, wherein each chimeric nucleic  
acid comprises an SMDP and/or SCP or functional fragment  
thereof.

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42. The method of claim 38 wherein said nucleic  
acid encoding a protein degradation binding domain is  
15 selected from the group consisting of Sia-1a, SIP-5,  
S, SAP-1, SAF-2, and SAD, or functional fragments  
thereof.

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43. A method for treating a disease by degrading  
the function of a target protein comprising:  
20 introducing, into a cell, a chimeric protein  
comprising a target-protein binding domain operatively  
linked to a protein-degradation binding domain of a  
protein member of the ubiquitin-mediated protein-  
degradation family.

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44. A chimeric protein comprising the SMDP and/or  
SCP of claim 13.

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SIP-L	MASEELOKDLEEVKVLLEKATRVRDALTAERSKIETEIRNNHQOKSOK	50
SIP-S	MASEELOKDLEEVKVLLEKATRVRDALTAERSKIETEIRNNHQOKSOK	50
SIP-L	KAEELDNEKEPAAVVAAPITTYJ2YKISNYGWDQSDKIVVVIYISTLTVHVP	100
SIP-S	KAEELDNEKEPAAVVAAPITTYJ2YDG1SQ1SL	80
SIP-L	TENVQVHFTTERGFOLLVNLNNGEYGSIVNNILFPISEVEGSSKKVKTOTV	150
SIP-S	-----	-----
SIP-L	LILCKXJVENTRHDYLTAQVEFECKEKEKPSYDTETOPSEGWLQSVLNKJIYE	200
SIP-S	-----	-----
SIP-L	DGDDDMKERTINKAHVEGREKQNKGDTEF	228
SIP-S	-----	-----

FIGURE 1

P	P
LYEDSGYSSFSL	SAD
SYLDSGIHSGAT	$\beta$ -catenin
DRHDSGLDSMKD	I $\kappa$ B $\alpha$
<u>DSG<math>\phi</math>XS</u>	
consensus	

FIGURE 2

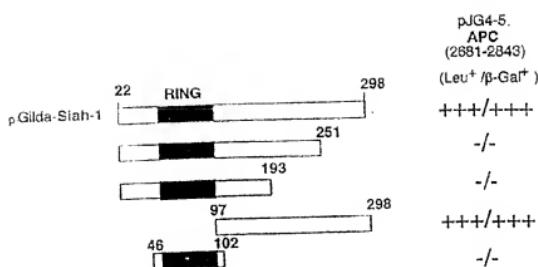


FIGURE 3

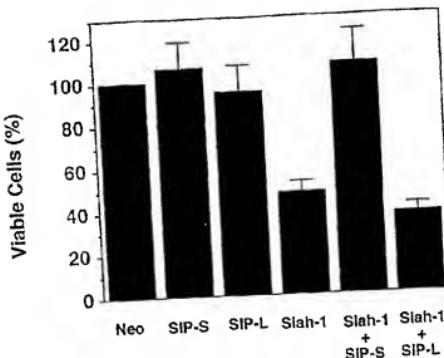


FIGURE 4

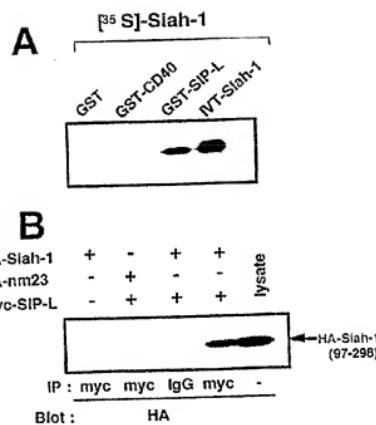


FIGURE 5

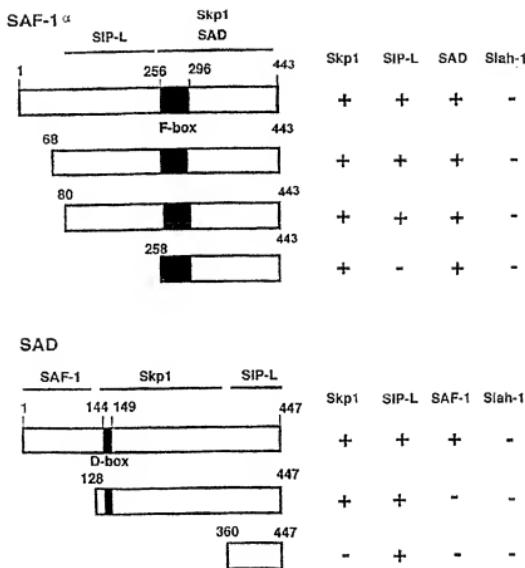


FIGURE 6

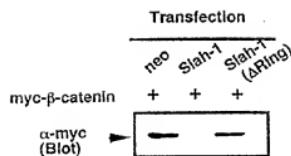


FIGURE 7

SIP: A novel E3 Complex Protein

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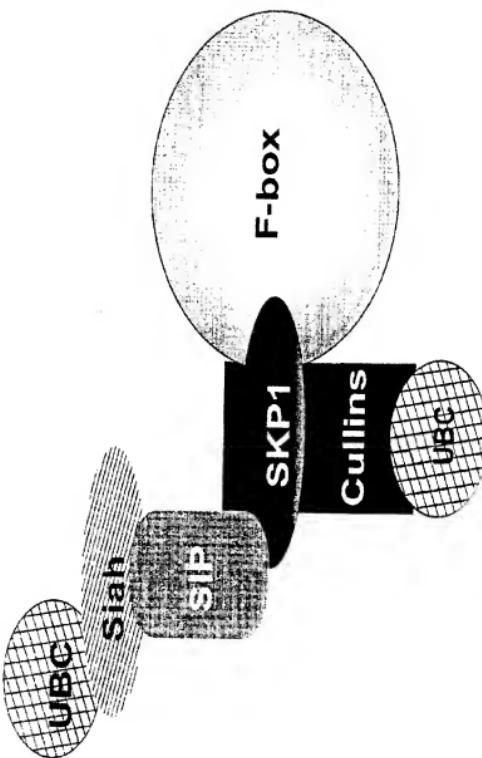


FIGURE 8

Scheme for Targeted Degradation of Endogenous TRAF Proteins Using  
SIP and TRAF-Binding Peptides

*Yeast two-hybrid peptide aptamer libraries*

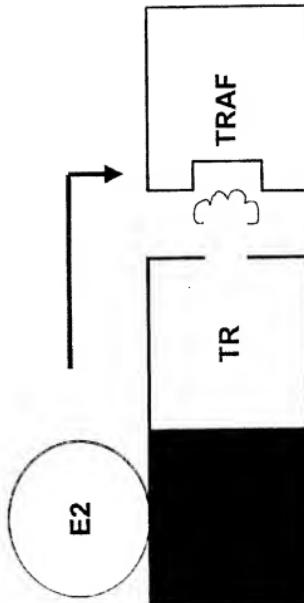


FIGURE 9

SIP Fused to TRAF-Binding Peptides induces Targeted Degradation of TRAF6  
**Yeast two-hybrid peptide aptamer libraries**

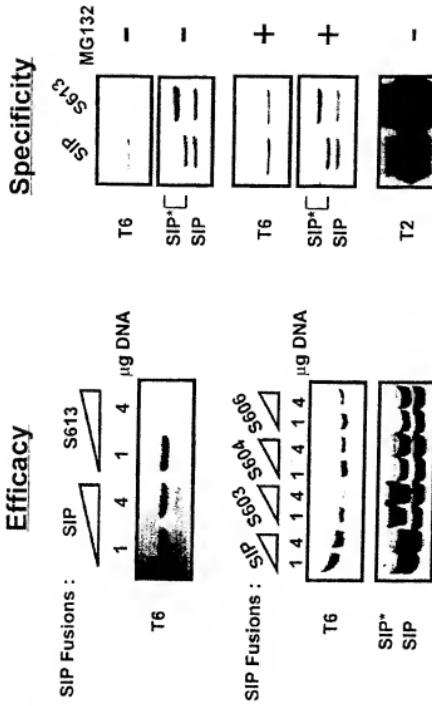


FIGURE 10

SEQUENCE LISTING

#### **2.1.6.3 The inward-looking economy**

c120. Nucleic Acid Encoding Proteins Involved  
in Protein Degradation, Products and Methods Related Thereto

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<150> US 09/330,517

$\times 1.53 \times 1.499 \times 0.6 = 1.2$

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 25          30           35

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Leu Glu Glu Val Lys Val Leu Ile Glu Lys Ala Thr Arg Lys Arg Val 99  
13                         15                         20                         25  
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30                         35                         40  
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aaa acc atg aat tat ggt tgg gat cag tca cat aag ttt gtg aac acc  
Lys Ile Ser Asn Tyr Gly Trp Asp Glu Ser Asp Lys Phe Val Lys Ile 291  
75                         80                         85  
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Tyr Ile Thr Leu Thr Val His Glu Val Val Pro Thr Glu Asn Val Glu 339  
90                         95                         100                         105  
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Val His Thr Glu Arg Ser Ilys Leu Val Lys Asn Leu Asn 387  
110                         115                         120  
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Gly Lys Ser Tyr Ser Met Cys Val Asn Asn Ile Lys Pro Ile Ser 435  
125                         130                         135  
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Val Glu Gly Ser Ser Lys Val Lys Asn Thr Asp Thr Val Leu Ile Leu 483  
140                         145                         150  
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Cys Arg Lys Dye Val Glu Asn Thr Arg Trp Asp Tyr Leu Thr Glu Val 531  
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Glu Lys Glu Cys Lys Glu Lys Glu Lys Pro Ser Tyr Asp Thr Glu Thr 579  
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gat cct aat gag gga ttg atg aat gtc cta aac aac att tat gag gat  
Asp Pro Ser Glu Gly Leu Met Asn Val Leu Lys Lys Ile Tyr Glu Asp 627







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 Glu Gly Cln Val Pro His Ser Leu Glu Thr Leu Tyr Glu Ser Ala Asp  
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 Cys Ser Asp Ala Asn Asp Ala Leu Ile Val Leu Ile His Leu Leu Met  
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 Leu Glu Ser Gly Tyr Ile Pro Gln Gly Thr Glu Ala Lys Ala Leu Ser  
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 Gly Asn Leu Ile Val Val Asn Ala Thr Leu Lys Ile Asn Asn Glu Ile  
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 Thr Pro Gln Gly Thr Glu Ala Lys Ala Ieu Ser Met Pro Glu Lys Trp  
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 Gly Ser Ser Ala Thr Leu Thr Cys Val Pro Leu Gly Asn Leu Ile Val  
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 275 280 285  
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 Pro Leu Leu Tyr Arg Phe Leu Tyr Leu Arg Asp Phe Arg Asp Asn Thr  
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 Asp Gln Arg Phe Thr Leu Pro Tyr Val Gly Asp Pro Ile Ser Ser Leu  
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 465 470 475 480  
 Arg Phe Asp Pro Val Gln Phe Leu Pro Gly Pro Asn Pro Ile Leu Pro  
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Lys Met Asp Leu Leu Ser Tyr Phe Gln Gln Leu Thr Phe Gln  
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Gln Ser Val Leu Leu Cys Gln Pro Glu Leu Glu Ser Ser Gln Ile  
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Asn Ile Ser Val Leu Pro Met Glu Val Leu Met Tyr Ile Phe Arg Trp  
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Val Val Ser Ser Asp Leu Arg Ser Leu Glu Gln Leu Ser Leu  
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Leu Ala Cys Leu Lys Val Trp Gly Arg Ser Cys Ile Lys Leu Val Pro  
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Tyr Ser Trp Arg Glu Met Phe Leu Glu Arg Pro Arg Val Arg Phe  
135 140 145  
gat ggg gtg eat atc aat gaa acc aca tcc aat aat cgt ccc ggg  
Asn Gln Val Tyr Ile Ser Ile Lys Thr Thr Tyr Ile Arg Gln Gly Glu Gln  
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Ser Leu Asp Glu Phe Tyr Arg Ala Trp His Gln Val Glu Tyr Tyr Arg  
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 Tyr Ile Arg Phe Phe Pro Asp Gly His Val Met Met Leu Thr Thr Pro  
 180 185 190  
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 Glu Gln Pro Gln Ser Ile Val Pro Arg Leu Arg Thr Arg Asn Thr Arg  
 195 200 205 210  
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 Thr Asp Ala Ile Leu Leu Gly His Tyr Arg Leu Ser Gln Asp Thr Asp  
 215 220 225  
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 245 250 255  
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 Asp Ser Lys Met Ala Asp Leu Leu Ser Tyr Phe Gln Gln Glu Leu Thr  
 35 40 45



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Val Lys Pro Asp Arg Ile Gly Arg Lys Val Ser Tyr Thr Pro Ala Tyr	
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Leu Glu Cys Ser Cys Lys Asp Cys Ile Lys Asp Tyr Glu Arg Leu Ser	
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Cys Ile Gly Ser Pro Ile Val Ser Pro Arg Ile Val Lys Leu Glu Thr	
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Glu Ser Lys Arg Leu His Asn Lys Glu Asn Gln His Val Glu Gln Thr	
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Leu Asn Ser Thr Asn Glu Ile Glu Asn Leu Glu Thr Ser Arg Leu Tyr	
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tcc tyc wtg cta cca ata cca aag cca gac gas tat ccc acc acc acc	639
Asn Cys Leu Leu Glu Ile Glu Ser Pro Asp Glu Tyr Pro Asn Lys Asn	
175	180
	185
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trg ctt gca gtt ctt cat ttt gaa aaw ggg gtt ttg tca aca tca aaa	667
Leu Leu Pro Val Ile His Phe Glu Lys Val Val Cys Asn Thr Leu Lys	
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Asn Asn Ala Lys Arg Asn Phe Lys Val Asp Arg Glu Met Leu Glu	
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Ile Ala Arg Gly Amn Phe Arg Leu Glu Asn Ile Ile Gly Arg Lys	
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Gly Leu Glu Cys Val Asp Ile Leu Ser Glu Leu Lys Asn Arg Gly	

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gtt gat aag ggg gca ttc cag ttg tac agt aac gca atc cas gaa gtt Asp Asp Lys Ala Phe Asn Leu Tyr Ser Lys Ala Ile Glu Arg Val 295	300	305	310	975
acc gaa aac aat aat aat ttt tat cat cat gct tca acc aya gaa tat Thr Glu Asn Asn Asn Lys Phe Ser Pro His Ala Ser The Arg Glu Tyr 316	320	325	330	1023
gtt atg ttt aca acc cca ctg got tat gtt cag aya tca gca gcc cag Val Met Phe Arg Thr Pro Leu Ala Ser Val Gln Lys Ser Ala Ala Glu 330	335	340	345	1071
act tat ctt aaa aaa gat got cas acc aag tta toc aat cca ggt gat Thr Ser Leu Lys Phe Asp Ala Gln Thr Lys Leu Ser Asn Gln Gly Asp 355	360	365	370	1119
cag aya gnt tat act tat agt cga cac aat gca ttc tct gag gtt gct Gln Lys Glu Ser Thr Tyr Ser Arg His Asn Glu Phe Ser Glu Val Ala 385	390	395	400	1167
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tgt tgc ttc ttt ttt Leu				1460



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 Leu Lys Lys Arg Ala Glu Thr Ile Leu Ser Asn Glu Gly Asp Glu Lys  
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 Gly Ser Thr Tyr Ser Arg His Asn Glu Phe Ser Glu Val Ala Lys Thr  
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 Leu Lys Lys Asn Glu Ser Leu Lys Ala Cys Ile Arg Cys Asn Ser Pro  
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 Ala Lys Tyr Asp Cys Tyr Leu Glu Arg Ala Thr Cys Lys Arg Glu Gly  
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 Cys Gly Phe Asp Tyr Cys Thr Lys Cys Leu Cys Asn Cys His Thr Thr  
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 Lys Arg Cys Ser Asp Gly Iys Leu Leu Lys Ala Ser Cys Lys Ile Gly  
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